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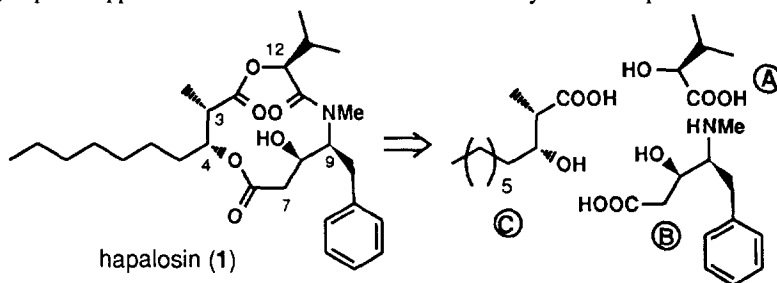
Synthetic Study on Hapalysin, a Cyclic Depsipeptide Possessing Multidrug Resistance Reversing Activities

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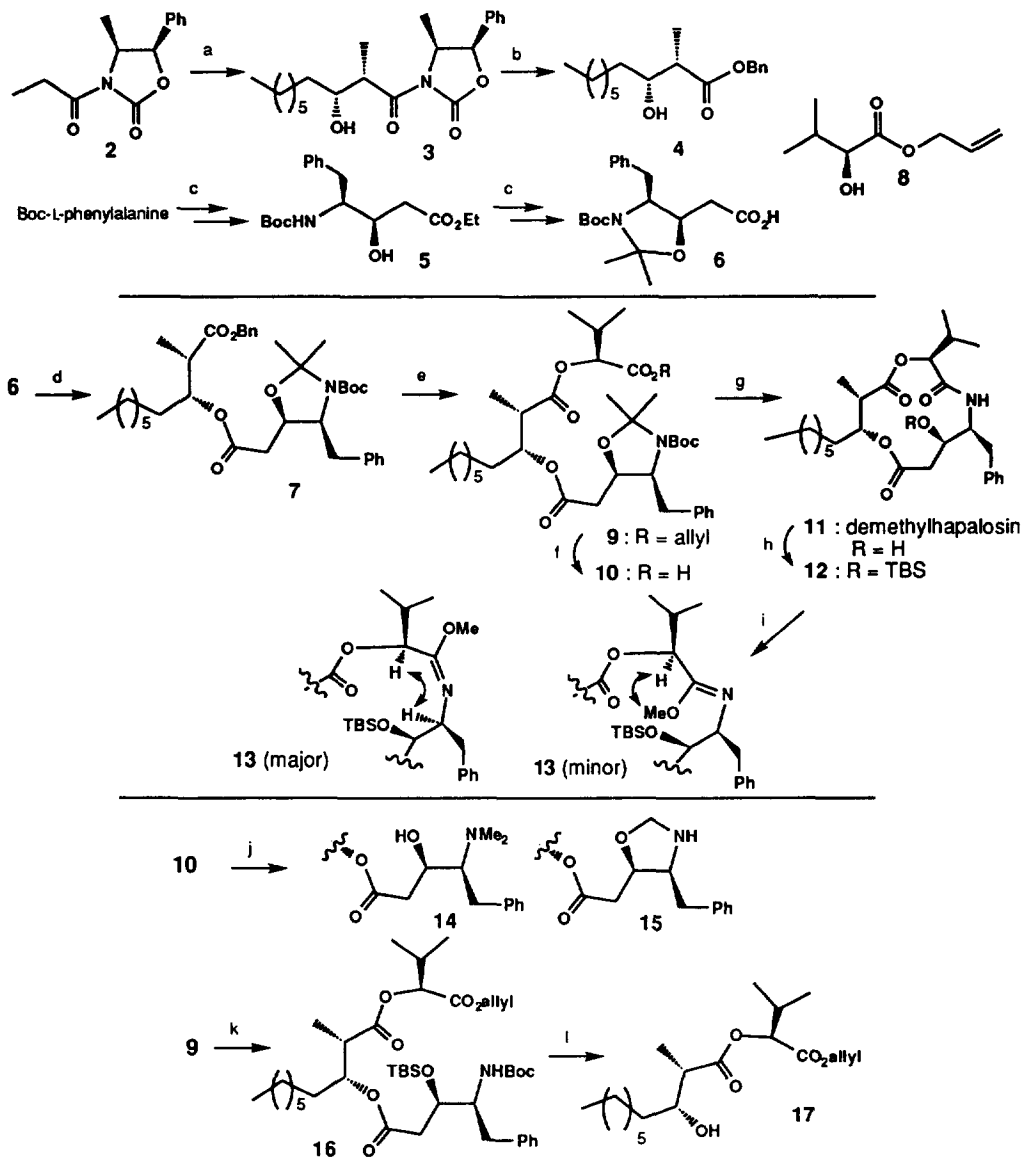
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Abstract: Hapalysin possessing a multidrug resistance reversing activity, has been synthesized from the corresponding hydroxy acids and γ -amino acids. The stereochemistry of the natural product and related derivatives is discussed. Copyright © 1996 Elsevier Science Ltd

Hapalysin (**1**), isolated from *Hapalosiphon welwitschii* W. & S. West, indicates better P-glycoprotein-mediated multidrug resistance reversing activity than verapamil to potentiate the cytotoxicities of such antitumor drugs transported by P-glycoprotein as daunomycin, vinblastine, actinomycin D, colchicine and taxol against resistant cells.¹ Among the many chemotherapeutic approaches to cancer disease, drug resistance remains one of the most urgent problems to be solved. Along with chemical interest in the cyclic depsipeptide structure, this background prompted us to initiate syntheses of **1** and its congeners.² When cleaved at the two esters and the amide bonds, the molecule was found to consist of 3-hydroxy-2-methylbutyric acid (**A**), 3-hydroxy- γ -amino acid (**B**) and β -hydroxy acid (**C**). In addition to commercially available **A** and the known structure (**B**),³ **C** might be prepared by the Evans aldol protocol.⁴ To enable the easy cyclization to the target molecule, the amide linkage would be finally constructed, and the following N-methylation might promise a facile introduction of diverse alkyl groups to supplement information on the structure-activity relationship.

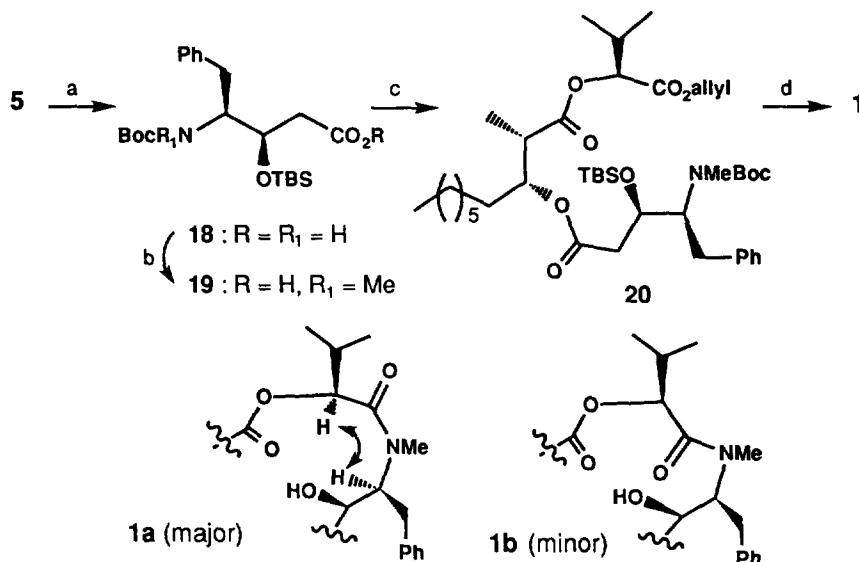


Oxazolidinone **2** was coupled with octanal in the presence of Bu_2BOTf and Et_3N to give the corresponding amide (**3**) in 90% yield, which on recrystallization contained no diastereomers detectable by the ^1H NMR spectrum. Treatment of **3** with LiOBn prepared from $n\text{BuLi}$ - BnOH provided the corresponding benzyl ester (**4**, 71%), which was condensed with the γ -amino acid (**6**)³ to give **7** in 86% yield based on **6**. Hydrogenolysis of **7** effected the abstraction of a benzyl group, leading to a carboxylic acid, which on coupling with **8** under the DCC - DMAP conditions furnished **9** in 81% yield in two steps. After removal of the protective groups of **9** by a 2-step procedure via **10**, the resulted acid underwent cyclization by a high dilution method (1 mmol/l) to produce demethylhapalysin (**11**).⁵ Transformation into the natural product (**1**) by



Scheme 1. a. Bu_2OTf , $\text{Et}_3\text{N}/\text{CH}_2\text{Cl}_2$, then $\text{Me}(\text{CH}_2)_6\text{CHO}$ (90%). b. $n\text{BuLi}$, BnOH/THF (71%). c. ref. 3. d. i) 2M NaOH : ii) **4**, DCC , $\text{DMAP}/\text{CH}_2\text{Cl}_2$ (86% from **6**). e. i) H_2 , $\text{Pd}(\text{OH})_2/\text{EtOH}$: ii) **8**, DCC , $\text{DMAP}/\text{CH}_2\text{Cl}_2$ (81% in two steps). f. $(\text{Ph}_3\text{P})_4\text{Pd}$, morpholine / THF (80%). g. i) $\text{TFA}/\text{CH}_2\text{Cl}_2$: ii) DPPA , $\text{EtNiPr}_2/\text{DMF}$ (1 mmol/l) (83% in two steps). h. TBSCl , Imd/DMF (92%). i. Me_3OBF_4 , Proton SpongeTM / CH_2Cl_2 (66%). j. i) $\text{TFA}/\text{CH}_2\text{Cl}_2$: ii) HCHO , $\text{NaBH}_3\text{CN}/\text{MeOH}$ (**14**, 72%): i) $\text{TFA}/\text{CH}_2\text{Cl}_2$: ii) HCHO , then NaBH_3CN (**15**, 67%). k. i) $p\text{-TsOH}/\text{MeOH}$ (67%): ii) TBSCl , Imd/DMF (82%). l. MeI , NaH/DMF (35%)

employing the protected derivative (**12**) was troublesome, probably owing to labile properties under the usual *N*-methylation conditions examined ($\text{MeI} - \text{NaH}$, $\text{MeI} - \text{Ag}_2\text{O}$ and $\text{MeI} - \text{LDA}$). During such attempts, exposure to Me_3OBF_4 and Proton SpongeTM,⁶ produced **13**,⁸ as a 1.8:1 (*cis* / *trans*) mixture of geometrical isomers, whose structures were deduced by the NOESY experiments, as depicted in Scheme 1. Interestingly, the



Scheme 2. a. i) TBSCl, Imd / DMF (100%); ii) 2M NaOH (84%). b. MeI, NaH / THF (77%). c. i) **4**, DCC, DMAP / CH₂Cl₂ (77%); ii) H₂, Pd(OH)₂ / EtOH; iii) **8**, DCC, DMAP / CH₂Cl₂ (86% in two steps). d. i) (Ph₃P)₄Pd, morpholine / THF (97%); ii) TFA / CH₂Cl₂ (94%); iii) DPPA, Et₃N / DMF (1 mmol/l) (44%).

preferential formation of the cis-type product is closely similar to the ratio of the natural conformers.¹ Additionally, N-methylation by using the primary amine was also unsuccessful; HCHO - NaBH₃CN gave a mixture of the dimethyl derivative (**14**), or acetal **15**. Upon employing **16** as a substrate, the MeI - NaH method gave rise to an ester cleavage to afford the fragment (**17**).

To circumvent the above-mentioned difficulties in the N-methylation, the N-methyl group was incorporated at an early stage of the synthesis. Thus, **5** was protected as a silyloxy ether, followed by hydrolysis under basic conditions to give **18** in 84% yield in two steps (Scheme 2). Compound **18** was submitted to MeI - NaH in THF⁹ to yield **19**. The following homologation reactions were performed by essentially the same procedure as mentioned above to give **20**. After removal of the protective groups, cyclization under high dilution conditions provided the target (**1**)¹⁰ in 44% yield. Spectral data of synthetic **1** were superimposable to those reported for the natural sample.¹

In particular, the ¹H NMR spectrum of synthetic **1** at room temperature showed a ~3:1 (**1a** / **1b**) mixture of the conformers as reported by Moore.¹ By further measurement (DMSO-d₆) at the elevated temperature, it was observed that the ratio of **1a** and **1b** reversibly changed to 1.7:1 (100 °C). Since NOE information was not sufficiently obtained, the conformation of the whole molecule could not be discussed. However, the NOESY experiments indicated that the major isomer (**1a**) exhibited a correlation between H₉ and H₁₂ ascribed to a cis-amide, while the minor (**1b**) adopted a trans-amide without a NOE effect at the same position.¹¹ Interestingly, contrary to the case of **1**, the ¹H NMR spectrum of demethylhapalosin (**11**) showed the presence of a trans-amide as a single isomer. Based on these observations, the N-methyl group may be a crucial factor to control the geometry of the amide bond.

Further investigation of the relationship between the N-alkyl group and the stereochemistry of the whole molecule, and concomitant biological activities are in progress.

References

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3. a) Maibaum, J.; Rich, D. H. *J. Org. Chem.* **1988**, *53*, 869 - 873. b) Parkes, K. E. B.; Bushnell, D. J.; Crackett, P. H.; Dunsdon, S. J.; Freeman, A. C.; Gunn, M. P.; Hopkins, R. A.; Lambert, R. W.; Martin, J. A.; Merrett, J. H.; Redshaw, S.; Spurden, W. C.; Thomas, G. J. *J. Org. Chem.* **1994**, *59*, 3656 - 3664.
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5. **11**: $[\alpha]_{\text{D}}^{22} -31.7^{\circ}$ (*c* 0.50, CHCl₃). Found *m/z* 475.2945. Calcd for C₂₇H₄₁NO₆: 475.2822 (M⁺). IR (film) 3320, 2940, 1735, 1665, 1595 cm⁻¹; ¹H NMR (400MHz, CDCl₃) δ 0.66 (3H, d, J= 6.8 Hz), 0.85 - 0.92 (6H, complex), 1.20 - 1.81 (16H, complex), 2.57 (1H, dd, J= 5.6, 14.0 Hz), 2.65 (1H, dd, J= 3.6, 14.0 Hz), 2.87 - 3.04 (3H, complex), 4.08 (1H, brs), 4.53 (1H, d, J= 8.0 Hz), 4.63 (1H, m), 5.46 (1H, d, J= 10.8 Hz), 5.52 (1H, m), 7.18 - 7.30 (5H, complex); ¹³C NMR (100.5 MHz, CDCl₃) δ 9.2, 14.1, 17.7, 18.5, 22.6, 25.5, 29.1, 29.3, 29.9, 31.0, 31.7, 37.5, 39.0, 41.2, 54.0, 70.6, 75.8, 81.8, 126.7, 128.6, 129.0, 136.9, 169.8, 173.4, 174.1.
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7. **12**: ¹H NMR (400MHz, CDCl₃) major conformer: δ 0.02 - 0.19 (6H, complex), 0.52 (3H, d, J= 6.8 Hz), 0.86 - 0.92 (6H, complex), 0.95 (9H, s), 1.17 (3H, d, J= 7.6 Hz), 1.20 - 1.49 (11H, complex), 1.95 - 2.05 (2H, complex), 2.30 (1H, dd, J= 10.0, 12.8 Hz), 2.54 (1H, dd, J= 5.6, 16.2 Hz), 2.76 (1H, dd, J= 2.4, 16.2 Hz), 2.87 (1H, m), 3.34 (1H, dd, J= 2.1, 12.8 Hz), 3.72 (1H, ddd, J= 2.1, 7.9, 10.0 Hz), 3.88 (1H, ddd, J= 2.4, 5.6, 7.9 Hz), 4.22 (1H, d, J= 8.8 Hz), 5.16 (1H, m), 7.08 - 7.31 (5H, complex).
8. Treatment with TBAF to remove a silyl protective group resulted in an inseparable mixture.
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10. **1**: $[\alpha]_{\text{D}}^{18} -41.0^{\circ}$ (*c*1.00, CH₂Cl₂). Found *m/z* 489.3079. Calcd for C₂₈H₄₃NO₆: 489.3090 (M⁺). IR (film) 3430, 2940, 1735, 1635cm⁻¹; ¹H NMR (400MHz, CDCl₃) major conformer: δ 0.23 (3H, d, J= 6 Hz), 0.57 (3H, d, J= 6 Hz), 0.88 - 0.92 (3H, complex), 1.14 - 1.40 (13H, complex), 1.50 - 2.05 (3H, complex), 2.61 (1H, m), 2.65 (1H, m), 2.86 (3H, m), 2.92 (1H, dd, J= 18.5 Hz), 3.22 (1H, m), 3.41 (1H, dd, J= 2.6, 14 Hz), 3.69 (1H, dt, J= 2.6, 10 Hz), 3.85 (1H, m) 4.31 (1H, d, J= 8.4 Hz), 5.12 (1H, m), 7.17 - 7.35 (5H, complex); ¹³C NMR (100.5 MHz, CDCl₃) major conformer: 12.1, 14.1, 17.5, 18.3, 22.6, 26.0, 28.0, 28.2, 28.8, 29.1, 29.2, 31.7, 36.4, 37.0, 40.7, 61.4, 70.2, 73.8, 76.5, 127.0, 128.9, 129.7, 137.4, 168.5, 168.7, 172.7.
11. The same observation was reported by Armstrong (see ref.2a).

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